Contents lists available at [ScienceDirect](http://www.sciencedirect.com/science/journal/09574166)

Tetrahedron: Asymmetry

journal homepage: www.elsevier.com/locate/tetasy

A nitro sugar mediated synthesis of 6-amino-1,5,6-trideoxy-1,5-imino-p-glucitol (6-amino-1,6-dideoxynojirimycin)

M. Begoña Pampín, Fernando Fernández, Juan C. Estévez *, Ramón J. Estévez *

Departamento de Química Orgánica, Universidade de Santiago de Compostela, 15782 Santiago de Compostela, Spain

article info

Article history: Received 27 January 2009 Accepted 24 February 2009 Available online 25 March 2009

ABSTRACT

6-Benzoylamino-3-O-benzyl-1,5,6-trideoxy-N-(1,1-diphenylmethyl)-1,5-imino-D-glucitol 12a and its epimer 6-benzoylamino-3-O-benzyl-1,5,6-trideoxy-N-(1,1-diphenylmethyl)-1,5-imino-L-iditol 12b, the protected forms of 6-amino-1,6-dideoxynojirimycin 4 and its C-5 epimer 5, respectively, were easily prepared from diacetone-p-glucose via 6-O-benzoyl-3-O-benzyl-1,2-O-isopropylidene-5-C-nitromethyl- α -pglucofuranose 7a or 6-O-benzoyl-3-O-benzyl-1,2-O-isopropylidene-5-C-nitromethyl- β -L-idofuranose 7b. 6-Amino-1,6-dideoxynojirimycin 4 (an important precursor of a wide range of glycosidase inhibitors) was easily prepared from 1,5-imino-D-glucitol 12a.

- 2009 Elsevier Ltd. All rights reserved.

Tetrahedron

1. Introduction

Glycobiology is an emerging research area in which glycosidases have received considerable attention on account of their involvement in a number of metabolic pathways.¹ Accordingly, the inhibition of glycosidases has become a powerful strategy for the treatment of several important diseases, such as diabetes, cancer and viral infections, including AIDS.² This situation has led to considerable effort in recent years aimed at understanding the structure types, action mechanisms and synthesis of glycosidase inhibitors.³

The most common glycosidase inhibitors are imino sugars such as the polyhydroxylated nitrogen-containing ring skeletons identical to those of sugars that mimic the enzyme. The parent iminosugar is nojirimycin 1 (Fig. 1), which can be regarded as the result of the replacement of the ring oxygen of p-glucose by a nitrogen atom. On the other hand, 1-deoxynojirimycin 2 (1-DNJ) and analogues are the compounds where the hemiacetal functionality is replaced by an aminomethylene group, a structural modification

that results in an improvement in the glycosidase inhibitory activity. 4 In fact, some of the latter compounds are therapeutically useful drugs.⁵

Our strategy was first applied to the D-glucose 6-amino derivatives of 1-DNJ, for example, compounds 4 and 5, which played an important role in glycosidase studies, not only because of their weak inhibition properties, but also because they are synthetic precursors of a wide range of glycosidase inhibitors.^{[6](#page-4-0)} Several methods have been described for the synthesis of 1-DNJ and its analogues[.4](#page-4-0) However, very few syntheses of 6-amino-1-deoxynojirimycins have been reported. $\frac{7}{2}$

Nitrosugars are powerful synthetic materials that combine the synthetic potential of sugars and the chemical versatility of the nitro group; for example, for the formation of carbon–carbon bonds prior to conversion into a range of other functionalities, including reduction to an amino group.^{[8](#page-4-0)} In connection with our recent investigations on nitrosugars, 9 we report here 9 we report here the new synthesis of 6-amino-1,6-dideoxynojirimycin 4, which is shown in [Scheme 1](#page-1-0).

Figure 1.

Corresponding authors. E-mail address: ramon.estevez@usc.es (R.J. Estévez).

^{0957-4166/\$ -} see front matter © 2009 Elsevier Ltd. All rights reserved. doi:10.1016/j.tetasy.2009.02.033

Scheme 1. Reagents and conditions: (i) KF·2H₂O, 18-crown-6 ether, CH₃NO₂, rt, 1 h (60% yield); (ii) H₂, Ni-Raney, MeOH, rt (52% yield); (iii) NaIO4, CH₂Cl₂, H₂O, SiO₂, rt, 6 h $(86\%$ yield); (iv) TFA, H₂O, rt, 12 h; (v) H₂NCHPh₂, NaCNBH₃, AcOH, MeOH, -78 to rt (63% yield); (vi) (a) Pd/C, HCOONH₄, 70 °C, 1 h; (b) HCl (6 M), 50 °C, 3 d (64% yield).

2. Results and discussion

A Henry reaction of the known p-glucose derivative ketone 6^{10} 6^{10} 6^{10} with nitromethane, using KF as the base, gave a 60% yield of a 58:42 mixture of the C-5 nitromethyl- α -D-glucofuranose 7a and the C-5 nitromethyl- β -L-idofuranose **7b**, as established by ¹H NMR spectroscopy. Careful chromatographic separation allowed the epimer 7a to be isolated and its structure was established by X-ray diffraction.¹¹

The ratio of 7a:7b can be explained in terms of the Felkin–Ahn model (Scheme 2). Thus, the depicted attack of the nitronate anion of nitromethane onto conformer 6a led to the major adduct 7a. On the other hand, the alternative attack of this nitronate anion on the carbonyl group of conformer **6b**, which is less favoured than the former, provided the minor stereoisomer 7b.

Hydrogenation of the mixture of $7a + 7b$ at room temperature, using Raney-Ni as the catalyst, unexpectedly provided a 52% yield of a 1:1.5 mixture of the p -gluco derivative 8a and the L -ido derivative $8b$, as established from the ${}^{1}H$ NMR spectrum of the epimeric mixture.

The formation of compound 8a can be explained in terms of the expected reduction of the nitro group of 7a to the amino group in compound 13 (Scheme 3), which under the reaction conditions rearranged spontaneously to 8a as a result of an intramolecular transfer of its benzoyl group from the oxygen to the nitrogen. Compound 8b should result from 7b in a similar manner.

Treatment of a suspension of the mixture of 8a + 8b and silica gel in dichloromethane with aqueous sodium periodate at room temperature resulted in an oxidative degradation of the 1,2-diole system, which gave ketone 9 in 86% yield. Compound 9 was then reacted with a mixture of trifluoroacetic acid and water in order to remove the acetonide protecting group. The resulting compound 10 was reacted with diphenylmethylamine in acidic conditions and underwent a heteroannulation process that provided the iminium

Scheme 3.

salt 11. This compound was reacted with sodium cyanoborohydride to give an epimeric mixture of $12a + 12b$, as a result of hydrogenation the $C=N$ bond of 11, from which both piperidines 12a (the thermodynamically more stable epimer) and 12b (the thermodynamically less stable component) were isolated in 24% and 9% yields, respectively, after careful column chromatography. The structure of the minor component 12b was established by X-ray diffraction (Fig. 2).^{[12](#page-4-0)}

Figure 2. Molecular structure of compound 12b in the solid state.

Finally, removal of the benzyl and benzoyl protecting groups of 12a, by treatment with $HCOONH₄$ as a hydrogen source and Pd/C as the catalyst followed by acidic hydrolysis with HCl (6 M), furnished the known six-membered iminosugar 4 (54% yield), which was isolated as its dihydrochloride salt.

3. Conclusion

In summary, we have described a new route to 6-amino-1 dideoxynojirimycins that is shorter than the rare previous examples. Work is now in progress to optimize this synthetic approach in order to apply it to hexoses other than D -glucose and thus to gain access to novel 6-amino-1-deoxynojirimycin analogues that could show improved activity profiles.

4. Experimental

Melting points were determined using a Kofler Thermogerate apparatus and are uncorrected. Specific rotations were recorded on a JASCO DIP-370 optical polarimeter. Infrared spectra were recorded on an MIDAC Prospect-IR spectrophotometer. Nuclear magnetic resonance spectra were recorded on a Bruker DPX-250 apparatus. Mass spectra were obtained on a Hewlett Packard 5988A mass spectrometer. Elemental analyses were obtained from the Elemental Analysis Service at the University of Santiago de Compostela. Thin layer chromatography (TLC) was performed using Merck GF-254 type 60 silica gel and ethyl acetate/hexane mixtures as eluants; the TLC spots were visualized with Hanessian mixture. Column chromatography was carried out using Merck type 9385 silica gel. Solvents were purified as per Ref. [13.](#page-4-0) Solutions of extracts in organic solvents were dried with anhydrous sodium sulfate.

4.1. 6-O-Benzoyl-3-O-benzyl-1,2-O-isopropylidene-5-Cnitromethyl-a-D-glucofuranose 7a and 6-O-benzoyl-3-Obenzyl-1,2-O-isopropylidene-5-C-nitromethyl-b-Lidofuranose 7b

 $KF\cdot 2H_2O$ (0.46 g, 4.90 mmol) and 18-crown-6 ether (0.82 g, 3.10 mmol) were added to a solution of ketone 6 (1.19 g) (2.90 mmol) in nitromethane (18 mL) and the resulting suspension was stirred at room temperature for 1 h. The reaction mixture was added to a mixture of ice/water (50 mL) and extracted with ethyl acetate (3×80 mL). The organic layers were then dried with anhydrous sodium sulfate, filtered and evaporated to give a residue, which was purified by flash column chromatography (ethyl acetate/hexane 1:3) to give 6-O-benzoyl-3-O-benzyl-1,2-O-isopropylidene-5-C-nitromethyl- α -D-glucofuranose **7a** (0.47 g, 34%) and 6-O-benzoyl-3-O-benzyl-1,2-O-isopropylidene-5-C-nitromethyl-b- L -idofuranose **7b** (0.36 g, 26%) as solids, which were crystallized from a mixture of ethyl acetate and hexane.

Compound $7a$: mp: 102-106 °C (ethyl acetate/hexane). $[\alpha]_D^{22} = -75.6$ (c 1.00, CHCl₃). IR (NaCl, v, cm⁻¹): 3454 (OH); 2854–3064 (C_{Ar}H); 1722 (CO); 1554, 1375 (NO₂). ¹H NMR (250 MHz, CDCl₃) δ 1.32 (s, 3H, CH₃); 1.46 (s, 3H, CH₃); 4.30 (d, 1H, $J_{3,4}$ = 3.35 Hz, H-3); 4.38 (d, 1H, $J_{4,3}$ = 3.35 Hz, H-4); 4.42-4.47 (m, 2H, CH₂NO₂); 4.54–4.60 (m, 2H, H-6 + H-6'); 4.66 (d, 1H, $J_{2,1}$ = 3.65 Hz, H-2); 4.73 (d, 1H, J = 11.87 Hz, CHPh); 4.80 (s, 1H, OH); 4.91 (d, 1H, $J = 11.87$ Hz, CHPh); 6.02 (d, 1H, $J_{1,2} = 3.65$ Hz, H-1); 7.31-7.61 (m, 8H, $8 \times$ HPh); 7.98-8.01 (m, 2H, 2 \times HPh). ¹³C NMR (62.8 MHz, CDCl₃) δ 26.17 (CH₃); 26.56 (CH₃); 65.34 (CH₂); 72.40 (CH₂); 73.05 (CH₂); 77.62 (CH); 77.76 (C); 81.38 (CH); 82.99 (CH); 104.53 (CH); 112.19 (C); 128.21 (2 \times C_{Ar}H); 128.45 (2 \times C_{Ar}H); 128.67 (C_{Ar}H); 128.79 (2 \times Ar C_{Ar}H); 129.06 (C_{Ar}) ; 129.54 $(2 \times C_{Ar}H)$; 133.39 $(C_{Ar}H)$; 135.47 (C_{Ar}) ; 165.64 (CO). MS (CI) m/z 474 [(M+H)⁺, 1]; 105 (38); 91 [(PhCH₂)⁺, 81], 28 (100).

Compound 7b: mp: $109-111$ °C (ethyl acetate/hexane). $[\alpha]_D^{22} = -38.0$ (c 1.90, CHCl₃). IR (NaCl, v_{max} , cm⁻¹): 3454 (OH); 2854–3089 (C_{Ar}H); 1722 (CO); 1554, 1375 (NO₂). ¹H NMR (250 MHz, CDCl₃) δ 1.33 (s, 3H, CH₃); 1.45 (s, 3H, CH₃); 4.27 (d, 1H, $J_{3,4} = 3.35$ Hz, H-3); 4.31 (s, 1H, OH); 4.39 (1H, d, $J_{4,3} =$ 3.35 Hz, H-4); 4.49-4.64 (m, 5H, H-6 + H-6' + CH₂NO₂ + CHPh); 4.69 (d, 1H, $J_{2,1}$ = 3.35 Hz, H-2); 4.74 (d, 1H, J = 11.8 Hz, CHPh); 6.01 (d, 1H, $J_{1,2}$ = 3.35 Hz, H-1); 7.33–7.60 (m, 8H, 8 \times H-Ph); 7.96–8.01 (m, 2H, 2 \times H-Ph). ¹³C NMR (62.8 MHz, CDCl₃) δ 26.60 (CH₃); 27.09 (CH₃); 65.90 (CH₂); 72.52 (CH₂); 73.90 (C); 78.96 (CH₂); 79.20 (CH); 81.80 (CH); 82.70 (CH); 104.90 (CH); 112.70 (C); 128.90 $(4 \times C_{Ar}H)$; 129.20 $(2 \times C_{Ar}H)$; 129.30 $(2 \times C_{Ar}H)$; 129.80 (C_{Ar}); 130.02 (C_{Ar}H); 133.70 (C_{Ar}H); 136.02 (C_{Ar}); 166.05 (CO). MS (CI) m/z 105 (38); 91 [(PhCH₂)⁺, 86], 61 (100); 28 (87).

4.2. 5-C-[(Benzoylamino)methyl]-3-O-benzyl-1,2-O-isopropylidene-a-D-idofuranose 8a and 5-C-[(benzoylamino)methyl]- 3-O-benzyl-1,2-O-isopropylidene-b-L-glucofuranose 8b

Raney-Ni (7.5 mL, 1 g/10 mL) was added to a deoxygenated solution of a 1:1 mixture of $7a + 7b$ (0.5 g, 1.05 mmol) in methanol (50 mL) and the resulting suspension was stirred at room temperature under a hydrogen atmosphere $(P = 1$ atm) for 15 h. The mixture was then filtered through Celite®, which was washed with methanol (50 mL), and the filtrate was evaporated to dryness. The residue was purified by flash column chromatography (ethyl acetate/hexane 2:1) to give 0.24 g (52%) of a 1:1.5 mixture of the epimers 5-C-[(benzoylamino)methyl]-3-O-benzyl-1,2-O-isopropylidene- α -D-glucofuranose 8a and 5-C-[(benzoylamino)methyl]-3-O-benzyl-1,2-O-isopropylidene-β-L-idofuranose 8b. IR (NaCl, v_{max} , cm⁻¹): 3378 (OH); 3064–2878 (C_{Ar}H); 1640 (CO). ¹H NMR (250 MHz, CDCl₃) δ 1.32 (s, 3H, CH_{3a}); 1.34 (s, 3H, CH_{3b});

1.47 (s, 3H, CH_{3a}); 1.49 (s, 3H, CH_{3b}); 3.09 (dd, 1H, J = 4.39 Hz, $J = 13.99$ Hz, CHNHBz_a); 3.25 (br s, 1H, OH_b); 3.33 (br s, 1H, OH_a); 3.46 (d, 2H, $J = 11.80$ Hz, CH_2OH_a); 3.56 (dd, 1H, $J = 4.39$ Hz, $J = 13.99$ Hz, CHNHBz_b); 3.66 (d, 2H, $J = 11.80$ Hz, CH₂OH_b); 4.01 (dd, 2H, J = 8.58 Hz, J = 13.99 Hz, 2 \times CHNHBz_(a+b)); 4.23 (d, 2H, $J_{3,4}$ = 3.029, 2 \times H_{3(a+b)}); 4.23 (d, 2H, $J_{4,3}$ = 3.029, 2 \times H_{4(a+b)}); 4.35 (s, 2H, 2 \times OH_(a+b)); 4.49–4.75 (m, 6H, 2 \times CH₂Ph_(a+b) + 2 \times H_{2(a+b)}); 5.99 (d, 1H, J_{12} = 3.84 Hz, H_{1a}); 6.03 (d, 1H, $J_{1,2}$ = 3.84 Hz, H_{1b}); 6.82–6.89 (m, 2H, $2 \times NHBz_{(a+b)}$); 7.27–7.79 (m, 16H, 16 \times H-Ph_(a+b)); 7.75–7.79 (m, 4H, $4 \times H$ -Ph_(a+b)). ¹³C NMR (62.8 MHz, CDCl₃) δ 25.87 (CH_{3a}); 25.94 (CH_{3b}); 26.34 (CH_{3a}); 26.36 (CH_{3b}); 43.33 (C_a); 43.37 (C_b); 63.12 (CH_{2a}); 63.75 (CH_{2b}); 71.69 (CH_{2a}); 71.86 (CH_{2b}); 74.91 (CH_{2a}); 75.19 (CH_{2b}); 78.85 (CH_a); 78.95 (CH_b); 81.10 (CH_a); 81.32 (CH_b); 82.80 (CH_a); 83.23 (CH_b); 103.98 (CH_a); 104.29 (CH_b); 111.51 (C_a); 111.53 (C_b); 126.86, 127.89, 127.95, 128.10, 128.20, 128.37, 128.44, 131.40, 131.46 (20 \times C_{Ar}H); 133.22 (C_{Ar-a}); 133.45 (C_{Ar-b}); 135.63 (C_{Ar-a}); 136.05 (C_{Ar-b}); 168.80 (CO_a); 169.25 (CO_b). MS (CI) m/z 444 [(M+H)⁺, 100]; 443 [(M)⁺, 5]; 442 [(M-H)⁺, 13]; 386 (63); 91 [(PhCH₂)⁺, 70].

4.3. 6-Benzoylamino-3-O-benzyl-6-deoxy-1,2-O-isopropylidene-a-D-xylo-hexofuran-5-one 9

A solution of a 1:1.5 mixture of the epimers $8a + 8b$ (0.23 g, 0.53 mmol) in dichloromethane (4 mL) was added to a suspension of silica gel (1.30 g), dichoromethane (2 mL) and a solution of NaIO₄ (0.23 g, 1.07 mmol) in H₂O (1.80 mL). The resulting mixture was stirred at room temperature for 6 h. The reaction mixture was filtered through Celite®, which was washed with dichloromethane (25 mL), and the filtrate was evaporated to dryness. The residue was purified by flash column chromatography (ethyl acetate/ hexane 1:2) to give 6-benzoylamine-3-O-benzyl-6-deoxy-1,2-Oisopropylidene- α -D-xylo-hexofuran-5-one 9 (0.19 g, 86%) as an oil. $[\alpha]_D^{22} = -94.8$ (c 1.20, CHCl₃). IR (NaCl, v_{max} , cm⁻¹): 3063– 2871 (C_{Ar}H); 1735 (CO); 1659 (CO). ¹H NMR (250 MHz, CDCl₃) δ 1.33 (s, 3H, CH₃); 1.48 (s, 3H, CH₃); 4.33 (d, 1H, $J_{3,4}$ = 3.51 Hz, H-3); 4.46 (d, 1H, $J = 11.72$ Hz, CHPh); 4.57-4.62 (m, 4H, $CH₂NHBz + CHPh + H-4$; 4.81 (d, 1H, $J_{2,1} = 3.51 Hz$, H-2); 6.11 (d, 1H, J_{12} = 3.51 Hz, H-1); 6.83–6.86 (m, 1H, NHBz); 7.17–7.31 (m, 5H, $5 \times$ H-Ph); 7.42–7.54 (m, 3H, $3 \times$ H-Ph); 7.79–7.82 (m, 2H, 2 × H-Ph). ¹³C NMR (62.8 MHz, CDCl₃) δ 26.14 (CH₃); 26.77 (CH₃); 49.03 (CH₂); 72.33 (CH₂); 81.55 (CH); 83.48 (CH); 84.57 (CH); 105.96 (CH); 112.43 (C); 126.94 ($2 \times C_{Ar}H$); 127.39 $(2 \times C_{Ar}H)$; 127.95 (C_{Ar}H); 128.37 (4 \times C_{Ar}H); 131.47 (C_{Ar}H); 133.82 (C_{Ar}); 136.46 (C_{Ar}); 166.95 (CO); 203.76 (CO). MS (CI) m/z 412 [(M+H)⁺, 100]; 354 (20); 91 [(PhCH₂)⁺, 87].

4.4. 6-Benzoylamino-3-O-benzyl-6-deoxy-a-D-xylo-hexofuran-5-one 10

Compound 9 (0.640 g, 1.55 mmol) was dissolved in a mixture $(1:1)$ of trifluoroacetic acid and H₂O (28 mL) and the resulting solution was stirred at room temperature for 12 h. The reaction was evaporated to dryness and coevaporated with toluene $(2 \times 1.5 \text{ mL})$ to give a yellow residue of 6-benzoylamine-3-O-benz y l-6-deoxy- α -D-xylo-hexofuran-5-one 10, which was used in the next step without further purification.

4.5. 6-Benzoylamino-3-O-benzyl-1,5,6-trideoxy-N-(1,1-diphenylmethyl)-1,5-imino-D-glucitol 12a and 6-benzoylamine-3- O-benzyl-1,5,6-trideoxy-N-(1,1-diphenylmethyl)-1,5-imino-Liditol 12b

A solution of compound 10 (570 mg) in methanol (11 mL) was added dropwise to a solution of diphenylmethylamine (1.22 mL, 1.24 mmol) and acetic acid (0.07 mL, 1.24 mmol) in methanol (11 mL) at -78 °C. The resulting reaction mixture was stirred at -78 °C for 30 min and sodium cyanoborohydride (0.2 g, 3.12 mmol) was then added in three portions over 45 min. The mixture was stirred for 2 h at -78 °C, allowed to warm up to room temperature and then stirred for 22 h. Saturated aqueous sodium bicarbonate (50 mL) was added and the mixture was extracted with dichloromethane (3×50 mL). The organic layers were dried with anhydrous sodium sulfate, filtered, evaporated in vacuo and the resulting residue was purified by flash column chromatography (ethyl acetate/hexane 1:2 to 1:1) to give 6-benzoylamine-3-O-benzyl-1,5,6-trideoxy-N-(1,1-diphenylmethyl)-1,5-imino-Dglucitol 12a (0.35 g, 44%) as an amorphous solid and 6-benzoylamine-3-O-benzyl-1,5,6-trideoxy-N-(1,1-diphenylmethyl)-1,5 imino-L-iditol 12b (0.15 g, 19%) as a yellow solid, which was crystallized from a mixture of ethyl acetate and hexane.

Compound **12a**: $[\alpha]_D^{22} = -1.3$ (c 0.50, CHCl₃). IR (NaCl, v_{max} cm⁻¹): 3365 (OH); 3063-2871 (C_{Ar}H); 1659 (CO). ¹H NMR (250 MHz, CDCl₃) δ 1.91 (dd, 1H, $J_{1,1'}$ = 11.52 Hz; $J_{1,2}$ = 10.15 Hz, H-1); 2.14 (br s, 2H, $2 \times OH$); 2.63–2.69 (m, 1H, H-5); 3.01 (dd, 1H, $J_{1,1}$ = 11.52 Hz; $J_{1,2}$ = 4.39 Hz, H-1'); 3.15 (t, 1H, $J_{3,4}$ = 8.50 Hz, H-3); 3.55 (t, 1H, $J_{4,3}$ = 8.50 Hz, H-4); 3.64–3.77 (m, 2H, H-6 + H-6'); 4.33-4.43 (m, 1H, H-2); 4.72 (d, 1H, $J = 11.52$ Hz, CHPh); 4.95 (d, 1H, $J = 11.52$ Hz, CHPh); 5.42 (s, 1H, CHPh₂); 6.56–6.61 (m, 1H, NHBz); 7.16–7.53 (m, 20H, 20 \times H-Ph). ¹³C NMR (62.8 MHz, CDCl₃) δ 36.95 (CH₂); 51.05 (CH₂); 62.17 (CH); 64.77 (CH); 69.28 (CH); 71.32 (CH); 74.57 (CH₂); 85.38 (CH); 126.72, 126.99, 127.04, 127.38, 127.60, 127.68, 127.89, 128.19, 128.36, 128.41, 129.96, 131.64 (20 \times C_{Ar}H); 133.48 (C_{Ar}); 137.05 (C_{Ar}); 138.43 (C_{Ar}); 141.39 (C_{Ar}); 168.80 (CO). MS (CI) m/z 523 [(M+H)⁺, 25]; 388 [(M-CH₂NHBz)⁺, 37]; 167 (100); 91 [(PhCH₂)⁺, 49].

Compound $12b$: mp: $128-130$ °C (ethyl acetate/hexane). $[\alpha]_D^{22} = +10.2$ (c 2.42, CHCl₃). IR (NaCl, v_{max} , cm⁻¹): 3355 (OH); 3084–2855 (C_{Ar}H); 1640 (CO). ¹H NMR (250 MHz, CDCl₃) δ 2.56 (dd, 1H, $J_{1,1'}$ = 11.52 Hz; $J_{1,2}$ = 10.15 Hz, H-1); 2.86–3.22 (m, 4H, $2 \times$ OH + H-1' + H-5); 3.52 (t, 1H, $J_{3,4}$ = 8.50 Hz, H-3); 3.61-4.02 (m, 4H, H-2 + H-4 + H-6 + H-6'); 4.78 (d, 1H, J = 11.52 Hz, CHPh); 4.90 (d, 1H, J = 11.52 Hz, CHPh); 4.95 (s, 1H, CHPh₂); 6.80-6.84 $(m, 1H, NHBz);$ 7.13–7.65 $(m, 20H, 20 \times H-Ph).$ ¹³C NMR $(62.8 \text{ MHz}, \text{CDCl}_3)$ δ 35.07 (CH₂); 48.97 (CH₂); 57.83 (CH); 70.16 (CH); 70.32 (CH); 72.05 (CH); 74.58 (CH₂); 83.54 (CH); 126.82, 127.15, 127.28, 127.36, 127.42, 127.84, 127.90, 128.45, 128.63, 128.73, 131.64 $(20 \times C_{Ar}H)$; 134.33 (C_{Ar}) ; 138.52 (C_{Ar}) ; 141.39 (C_{Ar}) ; 142.67 (C_{Ar}) ; 167.40 (CO). MS (CI) m/z 523 [(M+H)⁺, 99]; 388 [(M-CH₂NHBz)⁺, 100]; 167 (85); 91 [(PhCH₂)⁺, 68].

4.6. 6-Amino-1,5,6-trideoxy-1,5-imino-D-glucitol dihydrochloride 4

Pd/C $(10%)$ (185 mg) and HCOONH₄ $(60 \text{ mg}, 0.95 \text{ mmol})$ were added to a deoxygenated solution of compound $12a$ (70.5 mg, 0.134 mmol) in methanol (4 mL) and the resulting suspension was stirred at 70 \degree C under an argon atmosphere for 1 h. The reaction mixture was filtered through Celite®, which was washed with methanol (10 mL). The filtrate was concentrated to dryness and the residue dissolved in $H₂O$ (10 mL) and washed with dichloromethane (3×10 mL). The aqueous solution was evaporated to dryness, dissolved in aqueous HCl (6 M, 3 mL), stirred for 3 d at 50 \degree C and evaporated. The residue was dissolved in methanol and ethyl ether was added to give a white solid, which was filtered off and identified as 6-amino-1,5,6-trideoxy-1,5-imino-p-glucitol dihydrochloride 4 (17 mg, 54%). mp 187-188 °C (methanol/ethyl ether). $[\alpha]_D^{22} = +11.5$ (c 0.12, H₂O). IR (NaCl, v_{max} , cm⁻¹): 3425 (OH). ¹H NMR (250 MHz, CDCl₃) δ 3.92 (dd, 1H, $J_{1,1'} = 11.52$ Hz; $J_{1,2} = 12.11$ Hz, H-1); 4.25-4.49 (m, 6H, H-1' + H-3 + H-4 + H- $5 + H-6 + H-6$ '); 4.60–4.78 (m, 1H, H-2). ¹³C NMR (62.8 MHz, CDCl₃) δ 40.82 (CH₂); 47.97 (CH₂); 57.14 (CH); 68.18 (CH); 72.60 (CH);

76.95 (CH). HRMS: Calculated for $C_6H_{15}N_2O_3$ $(M+H)^+$ 163.1083; found 163.1095.

Acknowledgements

We thank the Spanish Ministry of Science and Innovation for financial support (CTQ2005-00555) and for a FPU grant to Fernando Fernández, and the University of Santiago de Compostela for a grant to Begoña Pampín.

References

- 1. (a) Winchester, B.; Fleet, G. W. J. Glycobiology 1992, 2, 199; (b) Look, G. C.; Fotsch, C. H.; Wong, C.-H. Acc. Chem. Res. 1993, 26, 182; (c) Nishimura, Y.. In Studies in Natural Products Chemistry; Atta-ur-Rahman, Ed.; Elsevier: Amsterdam, 1992; Vol. 10, pp 495–583; (d) de Raadt, A.; Eckhart, C. W.; Ebner, M.; Stütz, A. E. Top. Curr. Chem. 1997, 187, 157, and references cited therein.
- 2. (a) Martin, O. R.; Compain, P. Current Topics in Medicinal Chemistry; Bentham: Hilversum, The Netherlands, Vol. 3, Issue 5.; (b) Nash, R. J.; Watson, A. A.; Asano, N.. In Alkaloids Chemical and Biological Perspectives; Pelletier, S. W., Ed.; Elsevier Science Ltd: Oxford, 1996; Vol. 10, p 345.
- 3. Naoki, A.; Nash, R. J.; Russell, J. M.; Fleet, G. W. J. Tetrahedron: Asymmetry 2000, 11, 1645.
- 4. (a) Hughes, A. B.; Rudge, A. J. Nat. Prod. Rep. 1994, 11, 135; (b) Dilip, D. D.; Nabendu, N. S.; Vijaya, N. D. J. Org. Chem. 1997, 62, 7485; (c) Matos, C. R. R.; Lopes, R. S. C.; Lopes, C. C. Synthesis 1999, 4, 571; (d) Arnone, A.; Bravo, P.; Donadelli, A.; Resnati, G. Tetrahedron 1996, 52, 131; (e) Filtremann, J.; Duréault, A.; Depezay, J.-C. Synlett 1995, 235; (f) Johns, B. A.; Pan, Y. T.; Elbei, A. D.; Johnson, C. R. J. Am. Chem. Soc. 1997, 119, 4856.
- 5. (a) Butters, T. D.; Dwek, R. A.; Platt, F. M. Curr. Top. Med. Chem. 2003, 3, 561; (b) Platt, F. M.; Neises, G. R.; Dwek, R. A.; Butters, T. D. J. Biol. Chem. 1994, 269, 8362; (c) Block, X.; Lu, X.; Platt, F. M.; Foster, G. R.; Gerlich, W. H.; Blumberg, B. S.; Dwek, R. A. Proc. Natl. Acad. Sci. U.S.A. 1994, 91, 2235; (d) Mann, J.; Davidson, R. S.; Hobbs, J. B.; Banthorpe, D. V.; Harborne, J. B. Natural Products; Longman Scientific & Technical: England, 1994. p 442.
- 6. (a) McCort, I.; Fort, S.; Duéault, A.; Depezay, J.-C. Biorg. Med. Chem. 2000, 8, 135; (b) Heightman, T. D.; Vasella, A. Angew. Chem., Int. Ed. 1999, 38, 750.
- 7. (a) Tite, T.; Lallemand, M.-C.; Poupon, E.; Kunesch, N.; Tillequin, F.; Gravier-Pelletier, C.; Le Merrer, Y.; Husson, H.-P. Biorg. Med. Chem. 2004, 12, 5091; (b) Kilonda, A.; Compernolle, F.; Peeters, K.; Joly, G. J.; Toppet, S.; Hoornaert, G. Tetrahedron 2000, 56, 1005; (c) Ref. 6a.
- 8. (a) Barret, A. G. M.; Graboski, G. G. Chem. Rev. 1986, 86, 751; (b) Nitroalkanes and Nitroalkenes in Synthesis, Tetrahedron Symposia, Barret, A. G. M., Ed.; 1990; Vol. 46, p 7313.; (c) Noboru, O. In The Nitro Group in Organic Synthesis; Feuer, H., Ed.; Organic Nitro Chemistry Series; Wiley-VCH, 2001; (d) Luzzio, F. A. Tetrahedron 2001, 915.
- 9. (a) Otero, J. M.; Soengas, R. G.; Estévez, J. C.; Estévez, R. J.; Watkin, D.; Evinson, E. L.; Nash, R. J.; Fleet, G. W. J. Org. Lett. 2007, 9, 623; (b) Otero, J. M.; Fernández, F.; Estévez, J. C.; Estévez, R. J. Tetrahedron: Asymmetry 2005, 16, 4045; (c) Otero, J. M.; Barcia, J. C.; Estévez, J. C.; Estévez, R. J. Tetrahedron: Asymmetry 2005, 16, 11; (d) Soengas, R. G.; Pampín, M. B.; Estévez, J. C.; Estévez, R. J. Tetrahedron: Asymmetry 2005, 15, 211; (e) Soengas, R. G.; Estévez, J. C.; Estévez, R. J. Tetrahedron: Asymmetry 2003, 14, 3955; (f) Soengas, R. G.; Estévez, J. C.; Estévez, R. J. Org. Lett. 2003, 5, 4457; (g) Soengas, R. G.; Estévez, J. C.; Estévez, R. J.; Maestro, M. A. Tetrahedron: Asymmetry 2003, 14, 1653; (h) Soengas, R. G.; Estévez, J. C.; Estévez, R. J. Org. Lett. 2003, 5, 1423.
- 10. Fleet, G. W.; Smith, P. W. Tetrahedron 1987, 43, 971.
- 11. Valencia, L.; Estévez, J. C.; Estévez, R. J. Acta Crystallogr. 2009, E65, O332.
- 12. Crystallographic data for the structure of compound 11a have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication no. CCDC 715887. Copies of the data can be obtained free of charge on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK (fax (+44)1223-336-033; e-mail deposit@ccdc.cam.ac.uk).
- 13. Perrin, D. D.; Armarego, W. L. F. Purification of Laboratory Chemicals; Pergamon Press: New York, 1988.